



Global Advanced Research Journal of Agricultural Science (ISSN: 2315-5094) Vol. 5(7) pp. 302-308, July, 2016 Issue.
Available online <http://garj.org/garjas/home>
Copyright © 2016 Global Advanced Research Journals

Full Length Research Paper

Isolation and Identification of Fungi Associated With Tigernut Milk Drink (*Kunun Aya*) In Dutse, Jigawa State

Nura, M., Abubakar, A., Auyo, M, I., Sunday, E., and Kutama, A. S.

Department of Biological Sciences, Federal University, Dutse, Nigeria

Accepted 26 July, 2016

The study was carried out to isolate and identify the types of fungi associated with Tigernut milk Drink "*Kunun Aya*". The samples of kununaya were collected from five different areas within Dutse town. These include; Yalwawa, Fagoji, Gida dubu, Yan tifa, and Takur site. The experiment was carried out on fresh and spoilt samples, where the fresh samples were used for the first experiment, and the same samples were kept for three days to spoil and used for the second experiment. The serial dilution method was adopted for the experiments. The species of fungi isolated and identified were; *Aspergillus flavus*, *Aspergillus niger*, *Penicillium* species and *Saccharomyces cerevisiae*. The results showed that, the population of fungi were higher in spoilt samples than in the fresh ones.

Keywords: fungi, tigernut, Kunun Aya, Dutse

INTRODUCTION

Jigawa state is one of the 36 states in Nigeria, located in the north eastern part of Nigeria and it was founded in 27th of August, 1996. The capital of the state is Dutse. Jigawa state share border with Kano, Katsina, Bauchi, Yobe and Republic of Niger. Dutse is the capital city of Jigawa state. Dutse is situated at 11.76 north latitude, 9.34 East longitude and 460 meters elevation above the sea level. It is home to federal university Dutse which opened in November 2011. The estimated population of Dutse is 17,697 (2007). The research was carried out in Dutse town where some areas were randomly selected within Dutse metropolis. The areas are; Yalwawa, Fagoji, Gida dubu, Yan tifa and Takur site. The research was conducted on Tigernut milk drink (Tigernut milk drink) which is one of the popular local drink of the town.

Tigernut milk drink is a traditional non-alcoholic beverage with a spicy nutty taste. It is widely consume for its nutritional and medicinal benefits to its consumers and for its non-alcoholic properties (Abegas, 2007). It has immense social, economic, nutritional and medicinal benefits. Tigernut milk drink like other locally made drinks is widely consumed in Nigeria. In most Nigerians cities the sales and consumption of this locally made beverage is high. Due to the non-alcoholic nature of this drink, it is widely accepted and consumed as a substitute for alcoholic drinks. The drink is usually sold at the motor parks, school premises and market places and even served during social gatherings (Abegas, 2007)

In recent years, several developments in society have contributed to changes in the global beverage market. Consumers are increasingly aware of the impact of diet on their health and well-being. Beverages are not only consumed to provide refreshment and hydration, but also

*Corresponding Author's Email: kutamasak@yahoo.com

to increase well-being and to help in preventing nutrition-related disorders (Tenge and Geiger, 2001).

A range of microbes can be associated with drink manufacture, though only a few are able to cause spoilage. Microbiological spoilage leads to deterioration of the sensory quality and typically appears as off-flavors, odours and visual changes in the product (Stratford, 2006).

Tigernut milk drink is rich in nutrients (Abaejah *et al.*, 2006). It depends the internal mechanisms and prevents both constipation and diarrhea. Tiger nut has never been found to produce allergy (Belewa and Abodunrin, 2008). It is also used as a flavoring agent for ice cream and biscuits (Cantalejo, 1997).

Despite high nutritive value of this drink, its production in Nigeria has been hampered due to the deteriorating effects of some microorganisms on the drink (Abegas *et al.*, 2006). Osuntogun and Aboaba (2004) reported that Tigernut milk drink is prone to microbial deterioration due to either unhygienic methods of preparation or by use of contaminated raw materials and utensils (Osuntogun and Aboaba, 2004).

The aim of this research is the isolation and identification of common fungi associated with Tigernut milk drink.

Study Area

The study area was Dutse Metropolis, where the samples of Tigernut milk drink were obtained from randomly selected different areas within the town. The selected areas are: Yalwawa, Gida Dubu, Fagoji, Garu and Takur site in Dutse metropolis.

Sample Collection

Samples of freshly prepared Tigernut milk drink were obtained from randomly selected areas which include: Yalwawa, Gida Dubu, Fagoji, Garu and Takur site of Dutse town in Dutse local government Jigawa state. The samples were collected in 50cl plastics bottles. The bottles were labeled and taken to Biological science laboratory for analysis.

Preparation of Media

Two solid media potato dextrose agar (PDA) and sabouraud dextrose agar (SDA) were used for the isolation. The media were used in accordance with the manufacturers' instructions. A pure culture of each isolate was obtained by sub-culture, using the SDA plates.

Preparation of Sample

Two experiments were conducted, one for fresh, and the other for spoilt Tigernut milk drink sample. The fresh Tigernut milk drink samples were used for first experiment, the same samples were left for seven days to spoil.

Fivefold serial dilutions of each sample were prepared. Serial dilution was carried out, where 1ml of each sample was transferred into a test tube containing 9.0 ml of sterile distilled water and the test tube was shaken and labeled as 10^{-1} , from this tube 1.0ml was also transferred into another tube containing 9.0ml of the sterile distilled water and labeled as 10^{-2} . The procedure was repeated up to 10^{-5} using sterile syringes. The test tube 10^{-2} was used.

Inoculation

1.0ml from the dilution factors of each 10^{-2} test tube was transferred into sterile petri-dishes, containing prepared potato dextrose agar (PDA). The diluted samples were used to inoculate the prepared media using pour plate method. The agar plates were allowed to solidify and placed in an inverted position for 5 days at room temperature.

Isolation of Fungi

The samples of Tigernut milk drink were placed on potato dextrose agar (PDA) and incubated at room temperature for 5 days. After incubation, colonies of different shape and colors were observed on the plates (cheesbrough 2000). Pure culture of each colony type on each plate was obtained. This was done by sub-culturing each of the different colonies onto the SDA plates and incubated at room temperature again for 5 days Jiha (1995).

Identification

The identification of fungi was based on macroscopic and microscopic examination.

Macroscopic examination was based on color and nature of the hyphae.

In microscopic examination, the technique of James and Natalie (2001) was adopted for identification of unknown isolated fungi using cotton blue in lactophenol stain. The identification was achieved by placing a drop of the stain on clean slide, where a small portion of the mycelium was spread very well on the slide with the aid of the needle.

A cover slip was gently applied with little pressure to eliminate air bubbles. The slide was then mounted and observed with $\times 10$ and $\times 40$ objective lenses. The species encountered was identified in accordance with Cheesbrough (2002).

STATISTICAL ANALYSIS

The data were analyzed by using non parametric method of analysis. The method used was Kruskal-walles method (H test). The following table show the number of fungi in all the samples and their ranking.

Table 1: Identification table

S/N	Fungi species	Macroscopy	Microscopy
1	<i>Aspergillus flavus</i>	The upper surface of the colony was yellow- green with edge, granular surface and green coloration on the reverse side.	The conidiophores was thick walled, hyaline and slightly roughened, erect, long aseptate with a vesicle with short conidial chains.
2	<i>Aspergillus niger</i>	The colonies were widely spread black with smooth white edges and spongy surface densely packed and brown on the reverse side.	The conidiophore was long, erected from the base to the vesicle, smooth walled, hyaline with globes conidial head.
3	<i>Penicillium</i> species	Greenish or grey green colored colonies with greenish black color inside	Mycelium consist of highly branched network of septate hyphae. Many branched conidiophores sprout on the mycelium bearing conidiophore
4	<i>Saccharomyces cerevisiae</i>	The colonies were creamy or white in color.	Cells are large, globose and also budding.

Table 2: Name and Number of fungal isolate obtained in fresh samples

S/N	Species /Areas	Yalwawa	Gida dubu	Fagoji	Yan tifa	Takur site
1	<i>Aspergillus flavus</i>	5	5	6	6	6
2	<i>Aspergillus niger</i>	9	6	7	8	8
3	<i>Penicillium</i> species	6	8	6	7	7
4	<i>S. cerevisiae</i>	3	5	5	6	5
Total		23	24	24	27	26

RESULTS

Total of four fungal species *Aspergillus niger*, *Aspergillus flavus*, *Penicillium* species and *Saccharomyces cerevisiae* were isolated from five samples of Tigernut milk drink. They were identified by studying their macroscopic and microscopic characters (Table1) and compared with identification keys by Samson *et al.* (2004).

NUMBER OF COLONY

The following tables show the fungal species and the number of their colony of both fresh and spoilt samples of Tigernut milk drink found in Gida Dubu, Yalwawa, Fagoji, Takur Site and Yan Tifa.

After the finding the variance of each species of both fresh and spoilt samples, and the data do not met normality

assumption (constant variance). Non parametric method Kruskal-walles H-test were used.

By using Kruskal-walles H-test formular, -11.3 and -9.9 were calculated for both fresh and spoilt samples respectively. For K-1=4 degree of freedom at 0.05 significant level. From X^2 table $X^2_{0.95} = 9.49$ was obtained. Since -11.3 and -9.9 are less than 9.49, null hypotheses (H_0) was accepted and concluded that there was no significant difference on these fungi found in Tigernut milk drink samples of the areas.

DISCUSSION

All the five samples of Tigernut milk drink were found to be contaminated with the same number of fungal species. The presence of the fungal species in these samples indicate

Table 3: Name and Number of fungal isolate obtained in spoilt samples

S/N	Species /Areas	Yalwawa	Gida dubu	Fagoji	Yan tifa	Takur site
1	<i>Aspergillus flavus</i>	17	17	16	19	17
2	<i>Aspergillus niger</i>	15	19	20	24	23
3	<i>Penicillium</i> species	12	20	19	23	20
4	<i>S. cerevisae</i>	11	14	15	10	13
Total		55	70	70	76	73

Table 4: Number of fungi in fresh samples and their ranks

Yalwawa	R	Fagoji	R	Gida dubu	R	Yan tifa	R	Takur site	R
5	4	5	4	6	10	6	10	6	10
9	20	6	10	7	15	8	18	8	18
6	10	8	18	6	10	7	15	7	15
3	1	5	4	5	4	6	10	5	4
Total	35		36		39		53		47

(R= rank)

Table 5: Number of fungi in spoilt samples and their rank

Yalwawa	R	Fagoji	R	Gida dubu	R	Yan tifa	R	Takur site	R
17	10	16	8	17	10	19	13	17	10
15	6.5	20	16	19	13	24	19	23	18.5
12	3	19	13	20	16	23	18.5	20	16
11	2	15	6.5	14	5	10	1	13	4
Total	21.5		43.5		44		51.5		48.5

(R=rank)

poor hygienic method of preparation of Tigernut milk drink, unhygienic working environment or using unsterilized materials. Fungal isolates were identified by their cultural and morphological characteristics (Table 1). The species of

fungi found are *Aspergillus flavus*, *Aspergillus niger*, *Saccharomyces cerevisae*, and *Penicillium* spp. The study also indicate that spoilt Tigernut milk drink contain higher

percentage of fungi than fresh ones due to the increase in the population of the fungal species on storage.

In both fresh and spoilt samples, Sample collected from Yan tifa had the higher fungal population, and *Aspergillus niger* with the higher number of occurrence of 8 and 24 in both fresh and spoilt samples respectively. This indicate that, Yan tifa sample was prone to microbial contamination than the rest. And this also indicate the poor hygienic condition of the area. The occurrence of these fungi may cause diverse effects on human health as they have the potential of producing mycotoxins. The concentrations of these substances may increase during storage of Tignut milk drink due to the increase in the population of the fungal species, hence, daily intake of such Tignut milk drink containing mycotoxins could result in bioaccumulation in the body which could be hazardous to human health. *Aspergillus flavus* and *Aspergillus niger* are known to produce aflatoxins, ochratoxins which are carcinogenic and are capable of causing kidney and liver disorders, invasive and noninvasive aspergillosis, allergic and sinusitis (Bryce, 1999; Bennett and Klich, 2003 and Sumson *et al* (2004)

Aspergillus flavus strain produces two most common aflatoxins (B1 and B2) (Amaike *et al.*, 2011). *A. flavus* has a minimum growth temperature of 12c and a maximum growth temperature of 48c, the optimum growth temperature is 37c (Agrios and George 2005). The four major Aflatoxins produced by *A. flavus* are B1, B2, G1 and G2. The production of the major toxins are as a result of particular strains of *A. flavus*. Aflatoxin B1 is the most toxic compounds including aflatoxin, gliotoxin, kojic acid, aspartoxin and aspergillus acid (Hedayati, *et al.*, 2007). *A. flavus* aflatoxins can lead to acute hepatitis, immunosuppression, hepatocellular carcinoma and neutropenia in humans. It is highly possible for *A. flavus* to invade arteries of the lung or brain and cause infarction (Crawford *et al.*, 2005). *Aspergillus flavus* is the second leading cause of Aspergillosis (Crawford *et al.*, 2005).

Aspergillus niger's strains have been reported to produce potent mycotoxins called Ochratoxins (Abarca *et al.*, 1994). Recent evidence suggests some true *A. niger* strains do produce Ochratoxin A. it also produces the Isoflavoneorobol (Samson *et al.*, 2011). *A. niger* is less likely to cause human disease than some other *Aspergillus* species. Human may become ill but this is due to a serious lung disease Aspergillosis that can occur (Handwert and Brian, 2005). *Aspergillus niger* is one of the most common causes of Otomycosis (fungal ear infections), which can cause pain, temporary hearing loss, and in severe cases, damage to the ear canal and tympanic membranes (Handwert and Brian, 2005).

Some species of *Saccharomyces cerevisiae* are opportunistic pathogens that can cause infection in people with compromised immune systems (Cogliati, 2013). *S. cerevisiae* is not normally considered to be a pathogen. In healthy people, disease resulting from *S. cerevisiae*

colonizing in a particular area are very rare (Main, 1988). 1 percent of all vaginal yeast infections occur due to *S. cerevisiae* in the vagina, but symptoms associated with it are identical to the symptom caused due to another organism more commonly associated with yeast infections *Candida albican* (McCullough, 1988).

Penicillium glabrum produce citromycetin which is known to cause allergy, asthma and some respiratory problems (Cooley *et al.*, 2004, frisvad *et al.*, 1998).

REFERENCES

- Adeyemi T, Umar S (1994): Effect of manufacture of the quality characteristics of kunun zaki, a millet based beverage: *Niger food journal* 12:34-40
- Agrios GN (2005): 'Plant pathology' fifth edition. Elsevier Academic press p. 922.
- Ainsworth GC, Sparrow FK (1993): The fungi, Vol. 1 VA: London, Academic press pp:13- 67
- Amaha M, Kitabatake K (1981): Gushing in beer. In Pollock JRA (eds.) *Brewing Science*. Vol. 2. Academic Press, London, UK, pp. 457-489.104, pp. 334-338.
- Amaike S, Nancy P (2011): *Aspergillus Flavus*"Annual review of photopathology 49:107-133.
- Ayo JA (2004). Microbial evaluation of kununzaki and zoborodo drink locally produced and sold in a polytechnic community in Nigeria. *Food J.* 22:199-126.
- Back W (2005): Colour Atlas and Handbook of Beverage Biology. W. Back (ed.) Verlag Hans Carl: Nürnberg, Germany, p. 317.81 of new beverages. *Brewing Science*, Sept./Oct., pp. 128-134.
- Barnett HL, Hunter BB (1999): Illustrated genera of imperfect fungi. The Amer. Phytopathol. Soc. St. Paul Minnesota (USA); Aps. Press.
- Belew MA, Belew KY (2007): Comparative Physico-Chemical Evaluation of Tiger-nut, Soybean and Coconut Milk Sources. *International Journal of Agriculture Biology*, 9, 785-787.
- Boeira L, Bryce J, Stewart G, Flannigan B (1999a): Inhibitory effect of *Fusarium* mycotoxins on growth of brewing yeasts. 1 Zearalenone and Fumonisin B1. *Journal of the Institute of Brewing*, Vol. 105, pp.366-374.
- Burnett SL, Beuchat LR (2000): Human pathogens associated with raw produce and unpasteurized juices, and difficulties in decontamination. *Journal of Industrial Microbiology and Biotechnology*, Vol. 25, pp. 281-287. ISSN 13675435. doi: 10.1038/sj.jim.7000106.
- Cantalejo MJ (1997): Analysis of variable components derived from raw roasted earth almond (*Cyperus esculentus* L.) *J. Agric. Food chemistry* 45. 1853-1860.
- Cheesbrough M (2002): Biochemical tests to see bacteria in laboratory, practice in tropical countries, cheesbrough M (eds). Cambridge edn. Pp. 63-70.
- Cogliati M (2013): Global molecular epidemiology of *Cryptococcus neoformans*; *An atlas of the molecular types scientifica* 2013: 675213
- Crawford JM (2005): Liver and biliary tract pathogenic basic of disease, edn kumar V, *et al.*, 2005, Philadelphia; Elsevier sounders p. 924.
- Davenport RR (1996): Forensic microbiology for soft drinks business. Soft Drinks Management International, April 1996, pp. 34-35. Species in environments relevant to foods and beverages. *Journal of Applied Microbiology*, Vol. 96, pp. 1222-1229.
- Draeger M (1996): Physical observations on the subject of gushing. *Brauwelt International*, Vol. IV, pp. 363-367.
- Drusch S, Ragab W (2003): Mycotoxins in fruits, fruit juices and dried fruits. *Journal of Food Protection*, Vol. 66, pp. 1514-1527.
- Elmahmood AM, Doughari JH (2007): Microbial quality assessment of kunun zaki beverage sold in Gurei town Adamawa state *Afri-J. Food sci.* 11-15

- Engel G, Teuber D (2007). Food around the world: A cultural perspective. India: pearson Education pub. Ltd, New Delhi.
- Fairs A, Wardlaw, A.J, and Pashley, C.H. (2000). "guidelines on ambient intramural airbornefungalspores" *Journal of clinical immunology* 20(6): 490-98.
- Filtenborg O, Frisvad J (1996). Molds in food spoilage. *Int. J. Food microbial.* 2(4): 1-5.
- Filtenborg O, Frisvad J, Samson R (2004). Specific association of fungi to foods and influence of physical environmental factors. In *Introduction to food- and airborne fungi*
- Hammes W, Hertel C (2009). Genus I. *Lactobacillus beijerinckii* 1901, 212AL. In: DeVos, P., Garrity, G., Jones, D., Krieg, N. R., Ludwig, W., Rainey, F. A., Schleifer K.-H and Whitman, W. B. (edn.). *Bergey's Manual of Systematic Bacteriology*, 2nd edn. New York, USA: Springer. Pp. 465–513. ISBN 0-387-95041-9.
- Handwerk B (2005). Egypt's king Tut Curse, caused by Tomb Toxins; *National geographic*.
- Hedayati MT, Pasqualotto PA (2007). "Aspergillus flavus: human pathogen, allergen and mycotoxin producer" *microbiology* (153) : 1677- 1692
- Hobbs BC (1993). "Food poisoning and food hygiene" Edward Arnold.
- Hocking AD, Pitt JI (2001). Moulds. In Moir, C.J., Anderw-Kabilafkas, C., Arnold, G., Cox, B., Hocking, A.D. & Jensen, I. (eds). Spoilage of processed foods: causes and diagnosis. AIFST Inc. (NSW Branch) *Food Microbiology Group*, Australia, pp. 361–281.
- Hocking AD, Pitt JI (2015). Advances in food mycology. Springer
- Horsáková I, Voldich M, Šicnerová P, Ulbrich P (2009). *Asaiasp.* as a Bacterium Decaying the Packaged Still Fruit Beverages. *Czech Journal of Food Science, Special Issue*, Vol. 27, pp. S362–S365.
- Horwitz C (1981). *Diary microbiology*. Publ. National Dairy Council, London, pp: 71.
- James GC, Natalie S (2001). *Microbiology. A laboratory manual* (edn.) p 211-223
- Jiha DK (1995). *Laboratory manual on seed pathology*, vikas publishing house (PVT). Ltd. Pp. 13-30
- Kultura (2007). Proximate and Microbial analysis of burukutu and pito produced in Ilorin, Nigeria. *Afr. J. Biotechnol.* 6(5):587-590.
- Kendall P (2008). Microbiological and Nutritional qualities of dairy products: *Nat. Sci* 4(3) : 37-40.
- Laitila A, Alakomi HL, Raaska L, Mattila-Sandholm T, Haikara A (2002). Antifungal activities of two *Lactobacillus plantarum* strains against *Fusarium* moulds *in vitro* and in malting of barley. *Journal of Applied Microbiology*, Vol. 93, pp. 566– 576.
- Lawlor K, Schuman J, Simpson P, Taormina J (2009). In: Sperber, W.H. and Doyle, M.P. (eds.) Compendium of the Microbiological Spoilage of Foods and Beverages, *Food Microbiology and Safety*, pp. 245–283, Springer, New York.
- Main J, Mckenzie H, Paratte D (1988). Antibody to saccharomyces cerevisiae in Crohn's disease" *British medical journal*. Volume 297. P 1105 – 1106.
- Martorell P, Stratford M, Steels H, Fernández-Espinar MT, Querol A (2007). Physiological characterization of spoilage strains of *Zygosaccharomyces bailii* and *Zygosaccharomyces rouxii* isolated from high sugar environments. *International Journal of Food microbiology*, Vol. 114, pp. 234–242. ISSN 0168-1605.
- McCallum JL, Tsao R, Zhou T (2002). Factors affecting patulin production by *Penicillium expansum*. *Journal of Food Protection*, Vol. 65, pp. 1937–1942. ISSN0362-028X ER.
- Moore JE, McCalmont M, Xu J, Millar BC, Heaney N (2002a). *Asaiasp.*, an unusual spoilage organism of fruit-flavored bottled water. *Applied and Environmental Microbiology*, Vol. 68, pp. 4130–4131. ISSN 00992240. Doi: 10.1128/AEM.68.8.4130-4131.2002.
- Moore JE, Xu J, Heaney N, Millar BC (2002b). Spoilage of fruit-flavoured bottled water by *Gluconacetobactersacchari*. *Food Microbiology*, Vol. 19, pp 399–401. ISSN 0740-0020. doi: 10.1006/fmic.2002.0482.
- Moore-landecker (1996). *Basic food microbiology: 2ndedn*. The ohio state university, publishing press, Rein hold New york. Pp: 11-13.
- Mount, Michael (2007). Fungi and mycotoxins ' *vedmed.ucdavis.edu*.
- Munar M, Sebree B (1997). Gushing-a malster's view. *Journal of American Society of Brewing Chemists*, Vol. 55, pp. 119–122.
- Murphy P, Hendrich S, Landgren C, Bryant C (2006). Food Mycotoxins: an update. *Journal of Food Science*, Vol. 71, pp. 51–65.
- Osuntogun B, Abiola OO (2004): Microbiological and evaluation of some non- alcoholic beverages. *Pak. J. Nutr.* 3:188-192.
- Paterson RRM, Lima N (2010). Toxicology of mycotoxins. In: Luch A. (ed). *Molecular, clinical and environmental toxicology*. Clinical toxicology. Springer, Basel, Vol 2, pp 31–63.
- Pellaud J (2002). Gushing: state of the art. The Xth Jean de Clerck Chair, Leuven Belgium, 2002.
- Peter (1993). *Microbiology*, 5thedn. McGraw hill publishing company, New Delhi. P:272.
- Pitt J, Hocking, A.D. (1997). Fungi and Food Spoilage. In: Pitt, J.I. and Hocking, A.D. (eds.) 2nd edition, Blackie Academic & Professional, University Press Cambridge, UK.
- Pitt JI, Bacilico JC, Abarca ML (2000). "Mycotoxins and toxigenic fungi" p.41-46
- Raspor P, Goranovich D (2008). Biotechnological applications of acetic acid bacteria. *Critical Reviews in Biotechnology*, Vol. 28, pp. 101–124.
- Robert B (1974). Pathogenic fungi. Co. London, Toronto.
- Ross (1993). Nutrients composition and biological evaluation of Mesta seeds. *Plant food for human nutr.* 49:34-37.
- Samson R, Hoekstra E, Lund F, Filtenborg O, Frisvad, J. (2004). Methods for the detection, isolation and characterization of food-borne fungi. In *Introduction to food- and airborne fungi*. 7th ed. Samson, R., Hoekstra, E. & Frisvad, J. (eds). Centraal bureau voor schimmel cultures, Utrecht, The Netherlands, pp. 283–297.
- Sarlin T, Nakari-Setälä T, Linder, M., Penttilä, M. and Haikara, A. (2005). Fungal hydrophobins as predictors of the gushing activity of malt. *Journal of the Institute of Brewing*, pp. 111.
- Scholte R, Samson R, Dijkstrahuis J (2004). Spoilage fungi in the industrial processing of foods. In *Introduction to food- and airborne fungi*. 7th ed. Samson, R., Hoekstra, E. & Frisvad, J. (eds). Centraal bureau voor schimmel cultures, Utrecht, The Netherlands, pp. 339–359.
- Schuster E, Dunn-Coleman N, Frisvad JC (2002). On the safety of *Aspergillus Niger* – a review 59 (4-5): 426-435.
- Sperber WH (2009). Introduction to the, In: Sperber, W.H. and Doyle, M.P. (eds.) Compendium of the Microbiological Spoilage of Foods and Beverages, *Food Microbiology and Safety*, p. 1–39, Springer, New York.
- Stratford M (2006). Food and Beverage Spoilage Yeasts, In: Querol, H. and Fleet, G. (eds.) *Yeasts in Food and Beverages*, Berlin, Germany: Springer-Verlag, Chapter 11, pp. 335– 379.
- Stratford M, James SA (2003). Non-alcoholic beverages and yeasts. In: Boekhout, T. and Robert, V. (eds.) *Yeasts in Food*, Hamburg, Germany: B. Behr's Verlag GmbH & Co, Chapter 12, pp.309-345.
- Suzuki R, Zhang Y, Lino, T (2010). Novel acetic acid bacteria isolated from flowers in Japan. *Journal of General and Applied Microbiology*, Vol. 56, pp. 339–346.
- Tamplin M (2009). Predictive microbiology. In: Heredia, N., Wesley, I. and García, S. (eds.) *Microbiologically safe foods*. John Wiley & Sons, Inc. Pp 601– 610.
- Tenge C, Geiger (2001). Alternative functional beverages. *MBAA Tech. Quart*, Vol. 38, pp. 33–35.
- Teniola OD, Odanfa SA (2002). Microbial assessment and quality evaluation of "Ogi" during spoilage. *World J. microbial Biotechnol.* 18:731-737.
- The microbial world: yeasts and yeast-like fungi (2006) institute of cell and molecular Biology. retrieved 24 December 2006.
- Tribst AA, Sant'Ana S, de Massaguer PR (2009). Review: Microbiological quality and safety of fruit juices--past, present and future perspectives. *Critical Reviews in Microbiology*, Vol. 35, pp. 310-339. ISN 1549-7828; 1040-841X. doi: 10.3109/10408410903241428.
- Trickett, Jill (2001). The prevention of food poisoning. P: 8
- Tull, Anita. (1997). *Food and Nutrition 3rdedn*, Oxford university press p:154.

- Walker M, Phillips CA (2007). The growth of *Propionibacterium cyclohexanicum* in fruit juices and its survival following elevated temperature treatments. *Food Microbiology*, 6, Vol. 24, pp. 313–318. ISSN 0740-0020. doi: 10.1016/j.fm.2006.08.002.
- Walker M, Phillips CA (2008). The effect of preservatives on *P Alicyclobacillus acidoterrestris* and *Propionibacterium cyclohexanicum* in fruit juice. *Food Control*, Vol. 10, pp. 974–981. ISSN 0956-7135. doi10.10/j.foodcont.2007.10.003.
- Wareing P, Davenport DD (2005). Microbiology of soft drinks and fruit juices, In: Ashurst, P. (eds.) *Chemistry and Technology of Soft Drinks and Fruit Juices*, 2nd ed. Blackwell Publishing Ltd. Pp. 279–299.
- William CF (1998). *Food Microbiology*, 4thedn, publ. Tala McGraw Hill, New Delhi. Pp: 465-510. Abarca M, Bragulet M, Castella G, (1994). *Applied Environ World Gazetteer*. Stefan Helder. Retrieved 2007-02-18
- Yamada Y, Yukphan P (2008). Genera and species in acetic acid bacteria. *International Journal of Food Microbiol.*, 125(1), 15–24 microbial 60(7): 2650-2660.