



Global Advanced Research Journal of Agricultural Science (ISSN: 2315-5094) Vol. 8(3) pp. 109-113, March, 2019 Issue.
Available online <http://garj.org/garjas/home>
Copyright © 2019 Global Advanced Research Journals

Full Length Research Paper

Microalgae culture for the mosquito pre-adults feeding used as food in the aquaculture

¹Moises Esteban Mejia-Mejia, ^{2*}Elsah Arce Uribe, ³Judith García Rodríguez, ²José Figueroa Torres, ³Migdalia Díaz Vargas

¹Facultad de Ciencias Agropecuarias, Universidad Autónoma del Estado de Morelos, México.

²Laboratorio de Acuicultura, Centro de Investigaciones Biológicas, Universidad Autónoma del Estado de Morelos, México.
Tel: +52- 77- 73162354. orcid.org/0000-0002-9815-2525

³Laboratorio de Hidrobiología, Centro de Investigaciones Biológicas, Universidad Autónoma del Estado de Morelos, México.

Accepted 05 March, 2019

Microalgae are used as food in aquaculture because they have essential amino acids, polysaccharides, enzymes, and proteins, being a food that provides great benefits to cultures. Some mosquito pre-adults (*Culex quinquefasciatus*) are microalgae consumers and they are used as living food for fish. Mosquito pre-adults support various fish characteristics, for instance they retain the behaviour of hunters, fish develop more intense coloration, and increase their growth. We described the microalgae culture required for live food growth (mosquito pre-adults) in aquaculture, and recognized the amount of carotenoids encapsulate in these organisms that support the colour and immune system of fish consumers. We founded that *Chlorella* sp. is the most abundant microalgae in living food culture and this specie had a high carotenoid pigment concentration. Additionally, we founded that mosquito pre-adult keeps a high carotenoid pigment concentration (at initial and final culture cycle) which support that this living food is an excellent food to fish production.

Keywords: Microalgae abundance, living food, carotenoids, fertilization.

INTRODUCTION

Microalgae are microscopic organisms that usually lives suspended in the water column (Spolaore et al., 2006). In aquaculture, microalgae are used in nutrition and represent an important group because they have essential amino

acids, polysaccharides, enzymes, and proteins, being a food that provides great benefits to cultures (Muller-Feuga, 2000). Microalgae are used as fish food in the early development stages and as food for other groups that will later be supplied to the fish (Reitan et al., 1997). For instance, the microalgae, *Chlorella vulgaris*, it has been used to improve pigmentation in rainbow trout (Gouveia et al., 2002). Others organisms used as fish food are insect

*Corresponding Author's Email: elsah.arce@uaem.mx

larvae (Barroso et al., 2014; Luna-Figueroa et al., 2019). In several studies it has been found that the fish fed with mosquito larvae are more colourful and they are bigger in less time (Mares and Arce, 2017; Arce et al., 2018).

The characteristic that most positive influence has in the fish production is the nutritional quality food (Puello-Cruz et al., 2010). Quality food determines essential aspects for fish development such as the immune system, growth, coloration, and survival rate (Torrison, 1984; Yong and Lee, 1991; Clotfelter et al., 2007; Sefc et al., 2014). Commercial foods commonly in flakes or pellets presentation have consistently sought to improve each of these requirements; however, nowadays they are not always considered the best option for aquaculture (Sampaio et al., 2009). In view of this situation, live foods with high nutritional quality have been used (Woods, 2003). Mosquito pre-adults support various fish characteristics, among the most relevant are that the organisms fed with mosquito pre-adults retain the hunters behaviour, fish develop more intense coloration, and increase their growth (Katya et al., 2017). We described the microalgae culture required for live food growth (mosquito pre-adults) in aquaculture, and recognized the amount of present in these organisms that support the colour and immune system of fish consumers.

MATERIALS AND METHODS

Live food culture

The live food culture was prepared in 590 L tanks, the tanks were fertilized with one kilogram of chicken manure (8 tanks with a replica, total= 16 tanks, Nieves et al., 1996; Votolina et al., 1999). Once the was done, the tanks remained under observation and microalgae counts were carried out in order to know the abundance and the time collection of mosquito pre-adults, *Culex quinquefasciatus*. The temperature was registered daily during all culture cycle (HANNA HI98129 sensor).

Pigments measurement

In order to know the microalgae carotenoid pigments concentration, 100 ml of water was filtered from each culture live food tank with a Millipore filter (membrane pore opening 0.45 μm). Once the filtration was finished, the membrane was placed in a centrifuge tube with 10 ml of methanol and stirred until disintegrated. Subsequently, the sample was centrifuged for 10 minutes at 1500 rpm. The readings were made in a spectrophotometer (HACH DR/2010; De la Lanza and Hernández, 1998; Minh et al., 2014). The carotenoids concentration (mg/m^3) was recorded for each culture tank (16 tanks, at initial and final culture).

To obtain mosquito pre-adults pigments, mosquito pre-adults samples were collected from the culture tank. The samples were rinsed with water and macerated, which was dissolved with methanol (10 ml). Subsequently, the sample was centrifuged for 10 min at 1500 rpm; the supernatant fluid was placed in cells in order to be read in the spectrophotometer (HACH DR/2010; De la Lanza and Hernández, 1998 Minh et al., 2014).

The carotenoid pigments extraction was carried out twice per culture tank during the experiment period. The first extraction was performed on 7 day after fertilization because at thistime the coloration of the tank denotes microalgae presence. The second extraction was established considering that decreased and that the mosquito pre-adults presence was low.

Statistics

Carotenoids pigments concentration in a culture (microalgae and mosquito pre-adults) at initial and final culture cycle was evaluated using *t* test. All Statistical analyses were conducted using Statistics® 10 Software (Zar, 2010).

RESULTS

The most abundant microalgae species in the living food culture was *Chlorella* sp., (100 % at initial collection and 95 % at the final culture cycle). The microalgae abundance suitable for the living food culture and mosquito pre-adult collection was at 7 day next to the fertilization and it was 630431 ± 23922 cell/ml (Fig. 1-2). The final living food final cycle was when microalgae abundance was 226844 ± 10003 cel/ml (14 days next the fertilization, Fig. 1-2). The temperature culture was 27 ± 1.2 °C.

Mean microalgae carotenoids of the living food culture tank in all cycle were 1179 ± 200 mg/m^3 . Carotenoid concentration in microalgae was higher at initial living food collection (1750 ± 250 mg/m^3) than at the final collection (630 ± 150 mg/m^3 ; $t_{(2,8)} = 2.89$, $P < 0.05$; table 1). As to mosquito pre-adults mean carotenoids of the living food culture tank in all cycle were 587 ± 69 mg/m^3 . Carotenoid concentration in mosquito pre-adults was the same at initial living food collection (610 ± 100 mg/m^3) than at the final collection (570 ± 100 mg/m^3 ; $t_{(2,8)} = 0.006$, $P > 0.05$; table 1).

At the initial living food collection, carotenoids concentration was higher in microalgae (1750 ± 250 mg/m^3) than in the mosquito pre-adults (610 ± 100 mg/m^3 ; $t_{(2,8)} = 4.33$, $P < 0.01$; table 1) while at the final living food collection, carotenoids concentration was the same in microalgae (630 ± 150 mg/m^3) than in mosquito pre-adults (570 ± 100 mg/m^3 ; $t_{(2,8)} = 0.45$, $P > 0.05$; table 1).

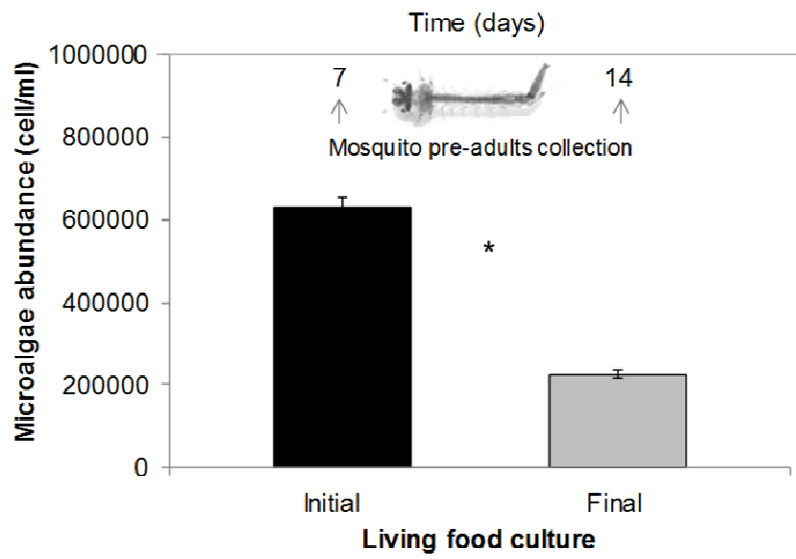


Figure 1. Microalgae abundance at the initial and final living food culture. Asterisks indicate significant differences ($P < 0.05$). Mosquito pre-adults collection period are shown.

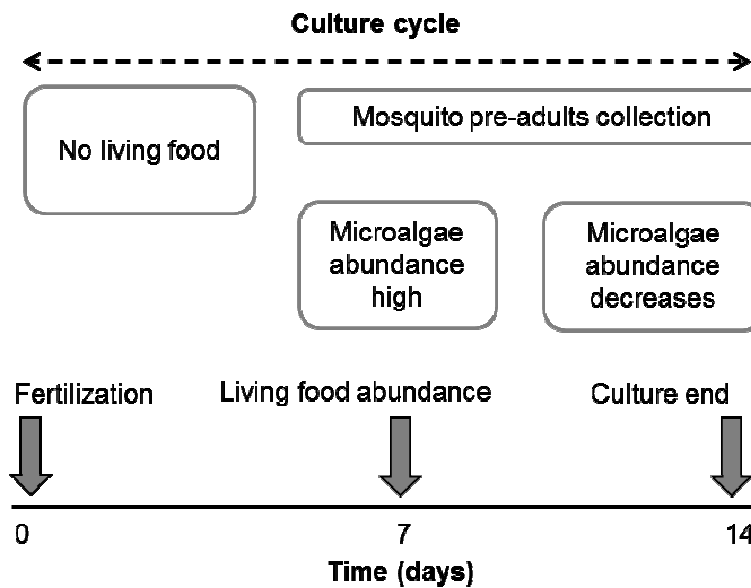


Figure 2. Microalgae living food culture cycle for feed mosquito pre-adults used as food in the aquaculture.

Table 1. Microalgae carotenoids pigments and mosquito pre-adults carotenoids pigments at the initial and final living food culture cycle. Mean values and SE are shown. Letters indicate significant differences ($P < 0.05$).

Organism	Carotenoids (mg/m ³)	
	Initial	Final
Microalgae	1750 ± 250 (a)	630 ± 150 (b)
Mosquito pre-adults	610 ± 100 (b)	570 ± 100(b)

DISCUSSION

Chlorella sp. is a common species in the microalgae culture (Gouveia et al., 2002). We founded that *Chlorella* is the microalgae that produces carotenoids pigment in our living food culture. Microalgae have high carotenoid concentrations, which are synthesized in the cells ribosomes and transported to chloroplasts (Yong and Lee, 1991). *Chlorella vulgaris* contains carotenoid pigments in concentrations of up to 0.4%, of which 80% are carotene (canthaxanthin and astaxanthin, Gouveia et al., 1996). The microalgae abundance determines the success of a live food culture since the zooplankton species (living food to fish) and other invertebrate larvae are feed with microalgae (Sipaúba-Tavares et al., 2016). The optimal microalgae abundance was presented 7 days after fertilization, at this time the carotenoids concentration is higher in microalgae than in mosquito pre-adults. Microalgae consumers are not able to produce carotenoids in a natural way; instead, they need to consume them from the primary producers as *Chlorella* sp. (Khatoon et al., 2010). Carotenoids expression in fish is considered an honest signal of the immune resistance (Houde and Torio, 1992). Additionally, carotenoids are responsible for the nutritional properties of fish muscle for human consumption (Rajasingh et al., 2006); while in ornamental fish the carotenoids determined the animal coloration and the price of this fish (Marañón et al., 1999). We founded that microalgae in culture has high carotenoids concentration. Gouveia and Rema (2005) observed the effect of microalgae biomass and the temperature in *Carassius auratus* on skin pigmentation. They founded that the best results were in fish fed with high levels of *C. vulgaris* in 26 to 30°C temperature range, and this range is

adequate for the microalgae culture (Paripatanamont et al., 1999). Additionally, *Chlorella* sp. increases the coloration of *Acipenser ruthenus* as complementary food (Sergejevova and Masojidek, 2013). We demonstrated that mosquito pre-adults are a food with a high concentration of carotenoids pigments that are made available to fish in production. Mosquito pre-adults does not undergo drying processes, freezing, or packaging, which decreases its original value, on the contrary, this food retains its nutritional value until ingestion and encourages consumers to hunt (Luna-Figueroa et al., 2009). Further, mosquito pre-adults obtain sources of carotenoids from microalgae present in cultured and even when the carotenoids pigments concentration of microalgae decreases, mosquito pre-adults are able to maintain the carotenoids pigment concentration until the end of the culture. Food fish has been enriched with microalgae, for instance, it has been used *Nostocellipsosporum* and *Navicula minima* to feed *Daphnia* sp. as food to Goldfish, *Carassius auratus* (Khatoon et al., 2010).

In conclusion, we founded that *Chlorella* sp. had a high carotenoid pigment concentration and we founded that mosquito pre-adult keeps a high carotenoid pigment concentration (at initial and final culture cycle) which support that this living food is an excellent food to fish production.

ACKNOWLEDGEMENTS

We thank Isela Molina (in memorial) for helpful comments on the original idea. We thank to Diana Molina, Yuritzi Castillo, Emmanuel Paniagua, and Marco Franco for the technical support in the laboratory experiments.

REFERENCES

- Arce UE, Archundia MPF, Luna-Figueroa J (2018). The effect of live food on the coloration and growth in guppy fish, *Poecilia reticulata*. *AS* 9: 171-179.
- Barroso FG, Haro C, Sánchez-Muros MJ, Venegas E, Martínez-Sánchez A, Pérez-Bañón C (2014). The potential of various insect species for use as food for fish. *Aquaculture* 422: 193-201.
- Clotfelter ED, Ardia DR, McGraw KJ (2007). Red fish, blue fish: trade-offs between pigmentation and immunity in *Betta splendens*. *Behav Ecol* 18: 1139-1145.
- De la Lanza EG, Hernández PS (1998). Nutrientes y productividad primaria en sistemas acuícolas. *Ecología de los sistemas acuícolas* 27-65 p.
- Gouveia L, Choubert G, Gomes E, Pereira N, Santinha J, Empis J (2002). Pigmentation of gilthead sea bream, *Sparus aurata* (L. 1875), using *Chlorella vulgaris* (Chlorophyta, Volvocales) microalga. *Aquacult Res* 33: 987-993.
- Gouveia L, Gomes E, Empis J (1996). Potential use of a microalgal (*Chlorella vulgaris*) in the colouring of rainbow trout (*Oncorhynchus mykiss*) muscle. *Lebensm Unters Forsch* 202: 75-79.
- Gouveia L, Rema P (2005). Effect of microalgal biomass concentration and temperature on ornamental goldfish (*Carassius auratus*) skin pigmentation. *Aquacult Nutr* 11: 19-23.
- Houde AE, Torio AJ (1992). Effect of parasitic infection on male colour pattern and female choice in guppies. *Behav Ecol* 3: 346-351.
- Katya K, Borsra MZS, Ganesan D, Kuppusamy G, Herriman M, Salter A, Azam SA (2017). Efficacy of insect larval meal to replace fish meal in juvenile barramundi, *Latescalcarifer* reared in freshwater. *IAR* 303-312.
- Khatoun N, Sengupta P, Homechaudhuri S, Pal R (2010). Evaluation of algae based feed in Goldfish (*Carassius auratus*) nutrition. *Proc Zool Soc* 63: 109-114.
- Luna-Figueroa J, Arce UE (2017). A diverse and nutritional menu in fish diets: "live feed". *Agroproductividad* 10: 112-116.
- Luna-Figueroa J, Arce UE, Figueroa TJ, Archundia FMP (2019). Pre-adults mosquito in fish species feeding. *IJAS* 10: 55-59.
- Marañón HS, Mayam PE, Salgado ZH (1999). Masculinización de *Xiphophorus helleri* (Pisces: Poeciliidae) inducida por los esteroides norgestrel y androstenediona. *Hidrobiología* 9: 31-38.
- Mares DOHA, Arce UE (2017). Live feed: coloration and growth rate in angel fish (*Pterophyllum scalare*). *Inv Agr* 14: 27-33.
- Minh AP, Hee-Guk B, Kyoung-Duck K, Sang-Min L (2014). Effects of dietary carotenoid source and level on growth, skin pigmentation, antioxidant activity and chemical composition of juvenile olive flounder *Paralichthys olivaceus*. *Aquaculture* 431: 65-72.
- Muller-Feuga A (2000). The role of microalgae in aquaculture: situation and trends. *J Appl Phycol* 12: 527-534.
- Nieves M, Voltolina D, Sapien MT, Gerhardus H, Robles AL, Villa MA (1996). Culturing microalgae with agricultural fertilizers. *Riv Ital Acquacolt* 31: 81-84.
- Paripatanamont T, Tangtrongpaioj J, Sailasuta A, Chansue N (1999). Effect of astaxanthin on the pigmentation of goldfish *Carassius auratus*. *J World Aquacult Soc* 30: 454-460.
- Puello-Cruz A, Velasco-Blanco G, Martínez-Rodríguez IE (2010). Growth and survival of Siamese fighting fish, *Betta splendens*, larvae at low salinity and with different diets. *J World Aquacult Soc* 41: 823-828.
- Rajasingh H, Øyehaug L, Vage DI, Omholt SW (2006). Carotenoid dynamics in Atlantic salmon. *BMC Biology* 4: 10.
- Reitan KI, Rainuzzo JR, Øie G, Olsen Y (1997). A review of the nutritional effects of algae in marine fish larvae. *Aquaculture* 155: 207-221.
- Sampaio ZJA, Salaro AL, Simões SMS, Moreno de Oliveira AL, Márcio BE, Siqueira AE (2009). Dietary protein and energy requirements of juvenile freshwater angelfish. *R Bras Zootec* 38: 989-993.
- Sefc KM, Brown AC, Clotfelter ED (2014). Carotenoid-based coloration in cichlid fishes. *Comp Biochem Physiol* 173: 42-51.
- Sergejevova M, Masojidek J (2013). Chlorella biomass as feed supplement for freshwater fish: sterlet, *Acipenser ruthenus*. *Aquacult Res* 44: 157-159.
- Sipaúba-Tavares LH, Appoloni AM, Fernandes JBK, Millan RN (2016). Feed of Siamese fighting fish, *Betta splendens*, (Regan, 1910) in open pond: live and formulated diets. *Braz J Biol* 76: 292-299.
- Spolaore P, Joannis-Cassan C, Duran E, Isambert A (2006). Commercial applications of microalgae. *J Biosci. Bioeng* 101: 87-96.
- Torrison OJ (1984). Effects of carotenoids in eggs and start-feeding on survival and growth rate. *Aquac Nutr* 43: 185-198.
- Votolina D, Nieves M, Piña P (1999). Fertilizers as cheap growth media for microalgae production: a Mexican point of view. *Riv Ital Acquacolt* 34: 43-45.
- Woods MCC (2003). Growth and survival of juvenile seahorse *Hippocampus abdominalis* reared on live, frozen and artificial foods. *Aquaculture* 220: 287-298.
- Yong YYR, Lee YK (1991). Do carotenoids play a photoprotective role in the cytoplasm of *Haematococcus lacustris*. *Phycologia* 30: 257-261.
- Zar JH (2010). *Biostatistical Analysis*. 5th Edition, Prentice Hall, Englewood Cliffs, NJ, 944 p.